

SARS-CoV-2 UV Dose Response Behavior

White paper prepared by IUVA COVID-19 Task Force

09/10/2020

Ernest R. Blatchley III, Lee A. Reith professor in environmental engineering, Purdue University

Brian Petri, research director, Trojan Technologies

Wenjun Sun, associate professor, Tsinghua University School of Environment

Ultraviolet (UV) radiation is a broad-spectrum antimicrobial agent that has been applied successfully in a wide range of disinfection applications. UV radiation in the wavelength range $200 \text{ nm} \lesssim \lambda \lesssim 320 \text{ nm}$, sometimes referred to as germicidal or microbicidal UV radiation, is known to cause damage to DNA and RNA that results in inactivation of microorganisms and viruses. For radiation with wavelengths less than about 240 nm, damage to proteins can also contribute to inactivation.

Given that all viruses contain a nucleic acid molecule, either DNA or RNA, and a protein coat (called a capsid) that surrounds the nucleic acid, all viruses are susceptible to inactivation by exposure to UV. However, viral sensitivity to UV is quite variable; the development of a comprehensive understanding of the causative factors is still an active area of research.

SARS-CoV-2 (aka the novel coronavirus) is the virus that causes coronavirus disease 2019 (COVID-19). Transmission of COVID-19 appears to be largely associated with airborne particles that may be released by symptomatic or asymptomatic individuals who have been infected by SARS-CoV-2,¹ although it also is known that the virus can remain infective on surfaces for as much as 24 to 72 hours, depending on the material it is in contact with,² so contact with surfaces represents another possible mechanism of disease transfer.³

At present, only limited data are available to define how SARS-CoV-2 responds to common disinfectants, including UV radiation. The standard method for defining the response of a microorganism to UV radiation is to measure its UV dose response behavior, which describes the kinetics of inactivation. UV dose response behavior represents a key piece of information for the design of UV disinfection systems.

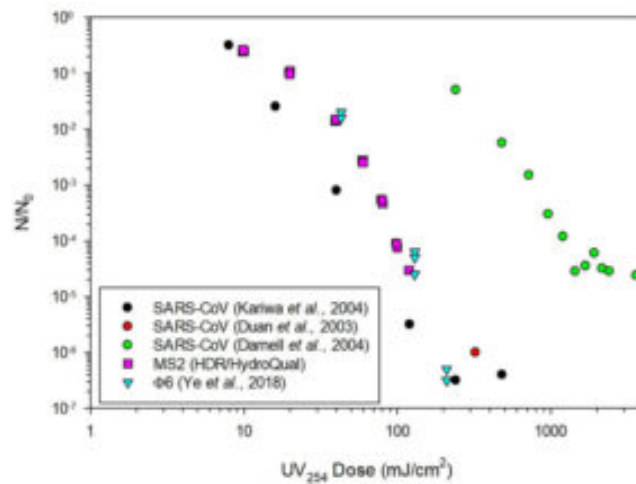


Figure 1. Reported UV254 dose response data for SARS-CoV in aqueous suspension.⁷⁻⁹ Note that the single data point provided by Duan et al. (2003) was based on radiation at 260 nm. Also included is a representative UV254 dose response data set for coliphage MS2 in aqueous suspension (data courtesy of HDR/HydroQual) and a dose response curve for Pseudomonas virus Φ6 from Ye et al. (2018)¹⁰.

Previous research has allowed definition of UV dose response behavior for other, closely related viruses. To date, three studies have been published that have reported UV dose response behavior of SARS-CoV (Figure 1); this is the virus that caused an epidemic of Severe Acute Respiratory Syndrome (SARS) that affected roughly 8,000 people in 26 countries in 2003.⁴ All of these studies were based on UV radiation at or near the wavelength (254 nm) that characterizes the output of low-pressure (LP) mercury lamps, which are the most commonly-used source of germicidal UV radiation.

Structurally, SARS-CoV and SARS-CoV-2 are similar viruses. Both are non-segmented, enveloped, single-stranded RNA (ssRNA) viruses. It is likely that the UV dose response behaviors of SARS-CoV and SARS-CoV-2 will be similar, though it is important to note that this has not yet been proven. Note also that there is considerable variability in the reported UV dose response behavior of SARS-CoV.

The variability evident in Figure 1 probably is due to deficiencies in the experimental methods that were used in some or all these previous studies. The stated fluences (doses) are probably overestimates, as the UV absorbance of the suspensions were not reported or explicitly accounted for. Similarly, it was not clear that the conditions of UV exposure would allow for accurate calculation of the UV dose applied to the viral suspension. Based on the information provided in the reported studies of SARS-CoV UV dose response behavior, the data set of Kariwa et al. appears to represent the most accurate, though it also may show false-high resistance.

Viruses often are characterized according to the presence/absence of an envelope, the form of their nucleic acid (single-stranded [ss] or double-stranded [ds]), and the type of nucleic acid (RNA or DNA); SARS-CoV-2 is an enveloped, ssRNA virus. The form of the genome may play an important role in governing the sensitivity of viruses to UV exposure.^{5,6} Emerging models based on nucleotide sequences have a strong basis with known UV disinfection mechanisms and may yield accurate predictions of the UV sensitivity of viruses.

Also included in Figure 1 are examples of the measured UV254 dose response behavior of coliphage MS2, a non-enveloped ssRNA virus, and Φ6, an enveloped double-stranded RNA (dsRNA) virus. These two viruses have similar UV sensitivity as the SARS-CoV of Kariwa et al. (2004). These data suggest that if the UV sensitivity of SARS-CoV-2 is similar to the Kariwa et al. (2004) data, MS2 and Φ6 may represent suitable non-pathogenic surrogates.

The effectiveness of UV-C radiation as a disinfectant is influenced by the wavelength of radiation. This behavior has become increasingly important in recent years with the development of alternatives to conventional LP or medium-pressure (MP) mercury lamps, including UV LEDs and plasma (excimer) lamps. The wavelengths of radiation produced by these sources depend on the chemical composition of the LED or the excimer. Collectively, these new sources provide access to radiation from across the germicidal UV spectrum.

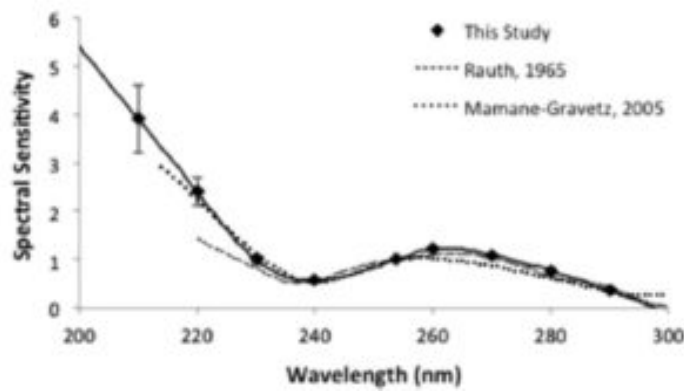


Figure 2. Normalized action spectrum for coliphage MS2 (figure from Beck et al., 2015).¹¹ For all three data sets presented in this graph, the data were normalized against the measured response at 254 nm. Also included in this figure are action spectra presented from Rauth (1965) and Mamane-Gravetz (2005).^{12,13}

A common graphical method for describing the effects of wavelength on microbial inactivation is the so-called action spectrum. In most cases, the action spectrum illustrates the relative rate of inactivation of a microbe or virus at a given wavelength compared to its inactivation rate in response to irradiation at 254 nm. An example of a normalized action spectrum for MS2 in aqueous suspension is presented in Figure 2. As with most action spectra, the information presented in Figure 2 indicates that for wavelengths in the range 240 to 300 nm, peak inactivation efficiency is observed at about 265 nm, with steady decreases at wavelengths above and below this peak. In this wavelength range, the majority of viral inactivation is attributable to photochemical damage to its nucleic acid. For radiation at wavelengths less than about 240 nm, a rapid increase in the efficiency of inactivation is observed; this is attributable to damage to proteins, which is known to take place at these short wavelengths.

It is expected that SARS-CoV-2 will display similar trends, but insufficient data are available at present to confirm or refute this hypothesis. The ultimate disinfection efficacy observed in an actual application will be influenced by the absorbance characteristics of the medium that is being disinfected. In some settings, there could be substances that could absorb strongly at wavelengths below 240 nm, which will mitigate the contributions of short wavelength UV-C radiation.

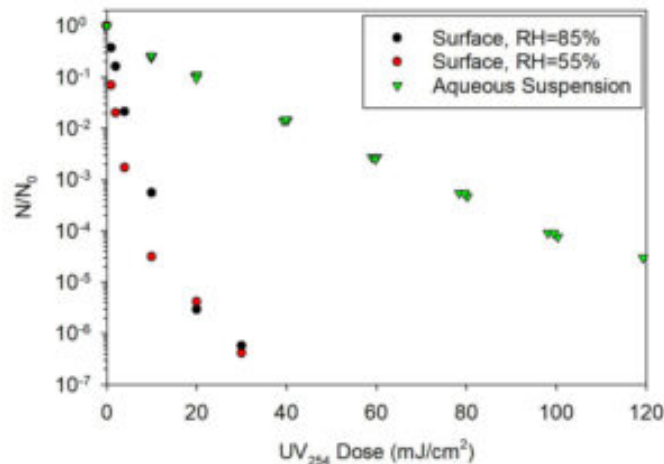


Figure 3. UV254 dose response behavior of coliphage MS2 on gel surfaces (at two different values of RH) and in aqueous suspension. Data for MS2 in surfaces from Tseng and Li (2007).¹⁸ Data for MS2 in aqueous suspension were provided by HDR/HydroQual.

Most UV dose response data have been reported for experiments wherein the target virus was suspended in water. These experiments are critical for UV disinfection of water; however, the mechanism of transfer of SARS-CoV-2 (and many other viruses) generally involves viruses that are suspended in air or attached to surfaces. For both conditions, the virus itself may not be suspended in water and may experience drying (desiccation). Desiccation, which will result from exposure to air

and will be influenced by relative humidity (RH), is known to represent a form of stress for most microbes and viruses and may alter their sensitivity to other forms of environmental stress, including UV exposure, as illustrated in Figure 3.¹⁴⁻¹⁸ However, it should be noted that some studies have indicated that similar viruses (including SARS and MERS) may survive on surfaces longer at high RH than at low RH.¹⁹

During the period of development of this white paper, several early reports of responses of SARS-CoV-2 and related viruses to UV radiation were reported in the scientific literature and in press releases. A study conducted by scientists at the US Department of Homeland Security indicated that UV-B radiation in ambient sunlight at irradiance values that are representative of mid-latitude, mid-day summertime conditions will result in 1-log₁₀ unit of inactivation in a time period of 7 to 15 minutes.²⁰ A preprint publication by Bianco et al. (2020) reported that a UV₂₅₄ dose of 3.7 mJ/cm² accomplished 3-log₁₀ units of SARS-CoV-2 inactivation when the virus was suspended in water.

A separate preprint document by Inagaki et al. indicated that a UV280 dose of roughly 38 mJ/cm² accomplished 3-log₁₀ units of inactivation of SARS-CoV-2. Buonanno et al. (2020) reported that far UV-C (i.e., wavelengths shorter than approximately 225 nm) accomplished at least 3-log₁₀ units of inactivation of aerosolized human coronaviruses alpha HCoV-229E and beta HCoV-OC43 at UV222 doses of 1.7 mJ/cm² and 1.2 mJ/cm², respectively.

Collectively, these results are promising; however, it should be noted that some important details of these experiments were not reported in these documents, specifically relating to methods used to quantify dose delivery to samples containing the target virus. It is likely publications in this area will continue to emerge, as well as the collective understanding of the effectiveness of germicidal UV radiation for control of SARS-CoV-2. To summarize, available evidence suggests UV radiation should be effective for inactivation of SARS-CoV-2 and other microbial pathogens. UV-based systems have important roles to play in battling SARS-CoV-2 in air, on surfaces and in water. However, like all common disinfectants (i.e., UV, chlorine, ozone), a need exists to quantify the kinetics of inactivation for SARS-CoV-2 for these applications.

Contact: Ernest R. Blatchley III, blatch@purdue.edu; Brian Petri, bpetri@trojanuv.com; Wenjun Sun, wsun@tsinghua.edu.cn

References

1. Prather, K. A.; Wang, C. C.; Schooley, R. T., Reducing transmission of SARS-CoV-2. *Science* 2020, 10.1126/science.abc6197.
2. van Doremalen, N.; Bushmaker, T.; Morris, D. H.; Holbrook, M. G.; Gamble, A.; Williamson, B. N.; Tamin, A.; Harcourt, J. L.; Thornburg, N. J.; Gerber, S. I.; Lloyd-Smith, J. O.; de Wit, E.; Munster, V. J., Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1. *New England Journal of Medicine* 2020, 382, (16), 1564-1567.
3. Marshall III, W. F. Can COVID-19 (coronavirus) spread through food, water, surfaces and pets? <https://www.mayoclinic.org/diseases-conditions/coronavirus/expert-answers/can-coronavirus-spread-food-water/faq-20485479>
4. WHO SARS (Severe Acute Respiratory Syndrome). <https://www.who.int/ith/diseases/sars/en/>
5. Qiao, Z.; Ye, Y. Y.; Chang, P. H.; Thirunarayanan, D.; Wigginton, K. R., Nucleic Acid Photolysis by UV254 and the Impact of Virus Encapsidation. *Environmental Science & Technology* 2018, 52, (18), 10408-10415.
6. Kowalski, W. J.; Bahnfleth, W. P.; Hernandez, M. T. In *A Genomic Model for Predicting the Ultraviolet Susceptibility of Viruses and Bacteria*, IUVA World Congress, Amsterdam, Netherlands, 2009; Amsterdam, Netherlands, 2009.
7. Kariwa, H.; Fujii, N.; Takashima, I., Inactivation of SARS coronavirus by means of povidone-iodine, physical conditions, and chemical reagents. *Japanese Journal of Veterinary Research* 2004, 52, (3), 105-112.

8. Duan, S. M.; Zhao, X. S.; Wen, R. F.; Huang, J. J.; Pi, G. H.; Zhang, S. X.; Han, J.; Bi, S. L.; Ruan, L.; Dong, X. P.; Team, S. R., Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. *Biomedical and Environmental Sciences* 2003, 16,(3), 246-255.
9. Darnell, M. E. R.; Subbarao, K.; Feinstone, S. M.; Taylor, D. R., Inactivation of the coronavirus that induces severe acute respiratory syndrome, SARS-CoV. *Journal of Virological Methods* 2004, 121,(1), 85-91.
10. Ye, Y. Y.; Chang, P. H.; Hartert, J.; Wigginton, K. R., Reactivity of Enveloped Virus Genome, Proteins, and Lipids with Free Chlorine and UV254. *Environmental Science & Technology* 2018, 52,(14), 7698-7708.
11. Beck, S. E.; Wright, H. B.; Hargy, T. M.; Larason, T. C.; Linden, K. G., Action spectra for validation of pathogen disinfection in medium-pressure ultraviolet (UV) systems. *Water Research* 2015, 70, 27-37.
12. Rauth, A. M., PHYSICAL STATE OF VIRAL NUCLEIC ACID AND SENSITIVITY OF VIRUSES TO ULTRAVIOLET LIGHT. *Biophysical Journal* 1965, 5, (3), 257-&.
13. Mamane-Gravetz, H.; Linden, K. G.; Cabaj, A.; Sommer, R., Spectral sensitivity of *Bacillus subtilis* spores and MS2 Coliphage for validation testing of ultraviolet reactors for water disinfection. *Environmental Science & Technology* 2005, 39, (20), 7845-7852.
14. Woo, M. H.; Grippin, A.; Anwar, D.; Smith, T.; Wu, C. Y.; Wander, J. D., Effects of Relative Humidity and Spraying Medium on UV Decontamination of Filters Loaded with Viral Aerosols. *Applied and Environmental Microbiology* 2012, 78 (16), 5781-5787.
15. Peccia, J.; Werth, H. M.; Miller, S.; Hernandez, M., Effects of relative humidity on the ultraviolet induced inactivation of airborne bacteria. *Aerosol Science and Technology* 2001, 35, (3), 728-740.
16. Fletcher, L. A., The Influence of Relative Humidity on the UV Susceptibility of Airborne Gram Negative Bacteria. *IUVA News* 2004, pp 12-19.
17. Ko, G.; First, M. W.; Burge, H. A., Influence of relative humidity on particle size and UV sensitivity of *Serratia marcescens* and *Mycobacterium bovis* BCG aerosols. *Tubercle and Lung Disease* 2000, 80, (4-5), 217-228.
18. Tseng, C. C.; Li, C. S., Inactivation of viruses on surfaces by ultraviolet germicidal irradiation. *Journal of Occupational and Environmental Hygiene* 2007, 4, (6), 400-405.
19. Otter, J. A.; Donskey, C.; Yezli, S.; Douthwaite, S.; Goldenberg, S. D.; Weber, D. J., Transmission of SARS and MERS coronaviruses and influenza virus in healthcare settings: the possible role of dry surface contamination. *Journal of Hospital Infection* 2016, 92, (3), 235-250.
20. Ratnesar-Shumate, S.; Williams, G.; Green, B.; Krause, M.; Holland, B.; Wood, S.; Bohannon, J.; Boydston, J.; Freeburger, D.; Hooper, I.; Beck, K.; Yeager, J.; Altamura, L. A.; Biryukov, J.; Yolitz, J.; Schuit, M.; Wahl, V.; Hevey, M.; Dabisch, P., Simulated Sunlight Rapidly Inactivates SARS-CoV-2 on Surfaces. *Journal of Infectious Diseases* 2020, XX, 1-9.
21. Buonanno, M.; Welch, D.; Shuryak, I.; Brenner, D. J., Far-UVC light (222 nm) efficiently and safely inactivates airborne human coronaviruses. *Scientific Reports* 2020.